Acoustic characteristics of microbubbles: Effect of acoustic pressure and pulse duration

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Laxman Subedi Biomedical Physics

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Abstract

The acoustic mechanism of ultrasound- Microbubble (MB) mediated sonoporation enhanced permeability of cell membranes has been associated with inertial and stable cavitations. The main objective of this study was to characterise acoustic behaviour of MB, under varying acoustic pressure and pulse duration using passive cavitation detection (PCD) technique. MBs were exposed to 1 MHz ultrasound pulse at varying acoustic pressure (0-1.5 MPa), pulse duration (5-30 μ s) at pulse repetition frequency of 1 KHz for insonation time of 1 second. The cavitation phenomena of MBs were detected passively using two transducers at center frequencies of 0.5 and 2.25 MHz. The MB acoustic response was characterised using FFT algorithm and cavitation dose method. Results indicated that the nonlinear oscillation of MBs increased with PNP and pulse duration. The integrated cavitation dose (ICD) increased with acoustic pressure and decreased with pulse duration. Definity MB showed greater ICD than Artenga MB.

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Dedication

To my Family

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List of Abbreviations

ACD	Active cavitation detection.
ART	Artenga.
DEF	Definity.
FDA	Food and drug administration.
FFT	Fast Fourier Transfer.
HMDEF	Harmonic for Definity.
HMART	Harmonic for Artenga.
ICD	Integrated cavitation dose.
MB	Microbubble.
PCD	Passive cavitation detection.
PNP	Peak negative pressure.
PRF	Pulse repitition frequency.
RMS	Root mean square.
RA	Receiver amplifier.
SHART	Subharmonic for Artenga.
SHDEF	Subharmonic for Definity.
TD	Time domain signal.
UCAs	Ultrasound contrast agents.
UH	Ultraharmonic.
UHART	Ultraharmonic for Artenga
UHDEF	Ultraharmonic for Definity.

Chapter 1

Introduction

1.1 Physics of Ultrasound

Ultrasound, sound at frequencies greater than 20 KHz, is a mechanical wave that propagates through a medium by the oscillation of its molecules or particles, transferring energy from one to another. In fluids and biological tissues, the oscillation of the particles is parallel to the direction of wave propagation where the propagating disturbances follow the form of pressure (longitudinal) waves with regions of compression (high density) and rarefaction (low density) (1). During propagation, the pressure waves interact with the underlying tissue resulting in scattering and absorption (2). Scattering of the ultrasound wave occurs when the pressure wave encounters regions of different acoustic impedances. The acoustic impedance of a material for a plane wave is given by

$$Z = \rho c, \tag{1.1}$$

Where ρ is the density of the medium and c is the speed of sound. The speed of

sound in liquids is given as

$$c = \sqrt{\frac{\beta}{\rho}} \tag{1.2}$$

where $(\beta = \rho \frac{dp}{d\rho})$ is the bulk modulus of the medium.

Moreover the scattering of ultrasound waves from a small spherical scatterer of diameter $a \ll \lambda$ at the far field depends on local variations in density and compressibility (3), given by equation

$$p_s(r,\Theta) = p_i e^{-jkr} \frac{k^2}{R^3} 3r[\frac{\kappa_\nu - \kappa_0}{\kappa_0} + \frac{3(\rho_\nu - \rho_0)}{2\rho_\nu + \rho_0} \cos\Theta]$$
(1.3)

where p_s is the scattered peak pressure, p_i is the incident pressure amplitude, $k = 2\pi f/c$ is the acoustic wave number and R = a/2 is the radius of the scatterer. Θ represents the angle between the incident wave and the radiated sound in dipole fashion and κ and ρ represent compressibility and density of the medium. The subscripts o and ν represents properties of the scatterer and surrounding medium respectively. The fractional change in compressibility given by the first term of the equation is the (monopole) term, which is the result of a pulsating point source while the difference in densities of the medium in the second term contribute scatterer to undergo back and forth motion, called as dipole term. In the case of air the scattering is mainly dominated by the compressibility term and thus air/gas-filled microbubbles have important applications as ultrasound contrast agents.

1.2 Ultrasound Contrast agents

Ultrasound contrast agents are comprised of shell-encapsulated gas-filled micron-sized (less than 10 μ m) bubbles, which are administered intravenously for imaging applications. From the clinical perspective microbubbles (MBs) have to be biocompatible, capable of crossing the pulmonary capillary bed after injection, and be stable enough to achieve contrast enhancement for the duration of ultrasound imaging. The first generation of MBs were air-filled bubbles without encapsulation which do not persist long enough to be of practical use in the human body. An improvement was made using higher molecular weight gases in a core and encapsulation with the shell. Gases such as perfluorocarbon, nitrogen or sulfur hexafluoride, have a lower Ostwald coefficient (the dimensionless ratio of solubility of the gas in liquid to gas density) and higher molecular weights which diffuse at a slower rate compared to those having a higher Ostwald's coefficient (4) as shown in Table 1.1.

Table 1.1:	Ostwald	coefficient	and di	sappearance	time (ti	me for	diffusion	of gas	when
microbubbl	le is in fre	ee gas state	e) for 3	μm diameter	: bubble	s having	g different	gases.	

gases	Ostwald's coefficient (10^6)	Disapperance $time(s)$
Air	23,168	0.02
Sulfur hexafluoride (SF_6)	5,950	0.1
Perfluropropane (C_3F_8)	538	1.1
Perflurohexane (C_6H_{14})	24	2

The encapsulation (shell) of the MB is comprised of protein, lipid or polymer layer. The shell affects the acoustic response of the MB, for example, the coating dampens the vibrations of the MBs and changes the microbubble resonance frequency (discussed in section 1.4).

1.3 Applications

MBs were originally developed to enhance the ultrasound scattered signals from blood vessels and the heart chamber. MBs are used clinically to improve imaging of cancerous tissues by detecting the tumor vasculature (5). In therapeutic applications, MBs can enhance the passage of bioactive materials across blood vessels (6), opening the blood brain barrier (BBB) to enhance local delivery of drugs to glioma (7). Moreover, MBs are used for the potential uptake of materials (drugs/genes) into cells or across biological membranes through transient opening of pores called sonoporation (8). In addition, MBs can be loaded with drug and genetic materials (9) and act as a vector of bioactive compounds. MBs can be tagged with ligands on their surface, and bind to specific disease sites in the body (10) that enhance targeted delivery.

1.3.1 Sonoporation

Sonoporation is a transient, reversible and nonlethal alteration of cell membrane permeability by ultrasound-MB mediated approach (11). This allows possible transportation of biologically active molecules such as DNA (12) chemotherapeutic drugs (13) genetic materials (14) and proteins delivery (15) to cells and tissues. Thus, sonoporation- mediated drug and gene therapy is currently being developed for cancer, and blood-brain-barrier limited treatments (16; 7). Figure 1.1 replicated from (11) illustrates how ultrasound and MB exposure enhance delivery of molecules across the cell membrane.

The MBs acoustic behaviour associated with sonoporation includes MB oscillation and disruption in an ultrasound field. Stable cavitation including linear and nonlinear oscillations of microbubbles (17; 18) and inertial cavitation (19) which is results of MBs disruption phenomenons have been shown to enhance cell permeability. Expanded



Figure 1.1: Sonoporation phenomenon: The application of ultrasound on MB increases the permeability of cell membranes and allows delivery of molecules into the cell.

knowledge of MB response may allow us to understand the biophysical mechanisms of snoporation and to improve and potentially control ultrasound-MB mediated therapeutic application.

1.4 MB Acoustic Behavior and Quantification

1.4.1 Microbubble acoustic behaviour

The acoustic behavior of the MBs depend on ultrasound exposure parameters such as acoustic pressure, frequency, pulse duration and Pulse repition frequency; and physical properties of microbubble such as size, concentration, gas and shell composition; and the surrounding environment (20). In the presence of the time-varying pressure of the US wave, MBs undergo a series of phenomena including formation, expansion or growth by rectified diffusion, and destruction of MBs called as acoustic cavitation (21). This process involves the conversion of low pressure of the ultrasound field into high pressure within small MBs (22). This acoustic energy within the MBs causes it to oscillate, expand and finally dissolve or collapse.

Cavitation commonly includes two types of bubble activities: namely, inertial and stable cavitations (Figure 1.2). Stable cavitation refers to prolonged volumetric shape oscillations of a MB, that can persist for several cycles in an ultrasound field (22). At lower acoustic pressure, the microbubble undergoes linear oscillation which is described by the linear oscillator model. The resonance frequency, ω_0 , of a free gas bubble, assuming a linear and an undamped system, (neglecting surface tension and damping) is given by

$$\omega_0 = \frac{1}{2\pi R_0} \sqrt{\frac{3\kappa_0 P_0}{\rho}} \tag{1.4}$$

where κ_0 represents the ratio of heat capacities at constant pressure to volume, R_0 is equilibrium radius of the MB, ρ is the density of the medium and P_0 is the static pressure of the surrounding medium at the bubble surface.

Equation (1.4) shows that the MB experiences a resonance that depends on the size of the bubble. It is believed when a bubble is driven at this resonant frequency, it reaches the highest oscillation amplitude and may provide more effective therapeutic results. In addition, for the case of encapsulated MB the shell increases the stiffness and viscous damping which dampens the motion of MB wall. This decrease the scattering ability of MB and thus increase the resonance frequency. At high acoustic pressures the microbubbles undergo nonlinear oscillation, which is described by modified Rayleigh Plessettee-type equation. Bubbles undergoing stable cavitation scatter the incident ultrasound beam with energy content at the driven frequency (f), during linear oscillations (expansion and contraction of MB by same amount) and harmonics (nf), subharmonics (f/2) or ultraharmonics (mf/n) during nonlinear oscillation (asymmetric expansion and



Figure 1.2: Microbubble Behaviours in an Ultrasound Field.

contraction) (23). This nonlinear oscillation has been explored for imaging applications. In addition MB oscillations in this regime produces circulating eddies (microstreaming) in the surrounding area of bubbles (24) with velocities and shear proportional to amplitude of oscillation, which is believed to cause stress on cell membranes and increase the permeability that enhance sonoporation.

Inertial cavitation is the rapid expansion (numbers of times the initial radius) and violent collapse (25) of the gas bubble that is caused at the higher acoustic pressure due to inertial effect of the surrounding liquid. It occurs for microseconds in few acoustic cycles which has limitations on the temporal resolution. Several mechanism of MB destructions including fragmentation into smaller bubbles, gradual gas diffusion and shell breakage are described previously. Destruction of MB is determined by coating of MBs. For rigid shelled MBs such as Quantison and BP127 the destruction is accompanied by shell rupture and gas release while soft shelled MBs such as Definity, Optison and Sonovue the disruption may be associated with violent collapse. MB undergoing this phenomenon scatters the acoustic energy in the form of broadband emissions related to post excitation signals (26). Moreover when the microbubble undergoes inertial collapse, this results in microjets (27) when it is close to the boundary that can puncture tissue and cell membranes, shockwaves (28) during implosion that produces stress on biological materials and free radical production have its application in sonoluminiscence. Since MBs collapse occurred suddenly with the large amount of acoustic energy released it is believed to cause lethal damage to biological tissues.

Beside these activities, At certain conditions of ultrasound exposures, the phospolipid coated microbubble exhibits complex phenomena such as buckling and ruptures (29) and compression only behaviours (30).

Stable and inertial cavitation mechanisms enhanced by MB activity play important roles in several ultrasound-based therapeutic applications such as shockwave lithotripsy to fragment kidney stones (31), thrombolysis (32; 33), tissue ablation (34), disruption of the blood brain barrier (BBB)(16) and targeted drug/gene delivery (35; 36; 37). In addition, cavitation has been studied for its significant potential in intracellular drug/gene delivery through the sonoporation technique, a medically significant approach that deliver the drugs/genes within the cells of the tissue in an efficient, spatially specific and safe way (8; 38; 17). Figure 1.3 replicated from (22) shows the different cavitation activities inducing bio-effects.

1.4.2 Effect of acoustic parameters on MB behaviour

As mentioned above the acoustic response of a MB depends on a number of factors, including its physical properties (e.g. size, gas and shell), ultrasound exposure parameters,



Figure 1.3: illustrate the cavitation mechanism inducing bio-effects.(A: therapeutic agent (triangles); B: gas bubble undergoing stable cavitation; C: microstreaming around cavitating bubble; D. collapse cavitation emitting a shock wave; E: asymmetrical bubble collapse producing a liquid jet that pierces the endothelial lining; F: completely pierced and ruptured cell; G: non-ruptured cells with increased membrane permeability due to insonation; H: cell with damaged membrane from microstreaming or shock wave; I: extravascular tissue; J: thin-walled microbubble decorated with agent on surface; K. thick-walled microbubble with agent in lipophilic phase; L: micelle with agent in lipophilic phase; M: liposome with agent in aqueous interior; N: vesicle decorated with targeting moieties attached to a specific target).

and environmental conditions (boundary condition, ambient pressure and temperature) (39; 40). Experiments have been carried out with varying acoustic pressure, frequency and pulse durations using acoustic methods. Experiments providing nonlinear oscillations of Optison MBs insonified at 2 and 4 MHz have shown that the first and second harmonic components increase linearly with acoustic pressure and the subharmonic and ultraharmonic components undergo rapid growth in the intermediate acoustic pressure

range (1.5MPa) (41). The MB acoustic behaviors of Optison and Definity were studied at peak to peak pressures (1.6, 2.0 and 2.4 MPa) with 2.8 MHz centre frequency transducer (42). It was reported that with increase of pressure the spectral magnitude increases for f/2, f/4 and for broadband spectra. The rupture threshold for Optison were studied at centre frequencies of 0.9, 2.8 and 4.6 MHz, pulse durations of 3, 5, 7 cycles of center frequency and peak negative pressures of 0.07 to 5.39 MPa. It was found that the rupture threshold increased with frequency and decreased with pulse duration (26). The stable and transient subharmonic emissions for Sonovue MBs, using 30 cycle pulse duration at 3.3 and 5 MHz and acoustic pressures of 40-225 kPa, was studied and have shown that emissions time of the stable and transient subharmonic thresholds were different (43). The subharmonic emission of 5 different agents at high frequency (25 MHz) have been investigated and have shown that with increase of pulse duration the subharmonic emissions are enhanced for all agents (44). The subharmonic emissions for Sonovue MB was studied at 3 and 30 cycle pulse duration and have shown that the emissions were higher at longer pulse duration of 30 cycles (45). Beside these the time dynamics of MBs at these varying acoustic conditions hasn't been investigated which is the focus of this thesis.

All of these finding motivate a rigorous investigation on the dependency of MB behavior on acoustic pressure and pulse duration. This may provide an understanding of MBs nonlinear oscillation with respect to time at exposure parameters which has important implication in inducing bioeffects.

1.4.3 MBs characterization: Passive Cavitation Detection (PCD)

Experimentally MBs have been characterized through acoustic emissions (25; 46) and optical imaging at high frame rates (47; 48; 30). Both of these techniques are employed for understanding the oscillation and disruption behaviour of the MBs. The acoustic method involves active cavitation detection (ACD) and passive cavitation detection (PCD) for sensing MBs behaviours. The ACD method is based on the pulse-echo technique, which measures the MB emissions generally with a single transducer. Since focus is the same for pulse and echo detection, with the same transducer, this method has greater spatial and temporal resolution (49). In PCD, the transducers or hydrophones are operated in receiving mode only, where the receiver measures the emissions from MB (49; 50; 51; 52). In this study we measure the bubble emissions using the passive cavitation detection (PCD) technique.

1.4.4 Microbubble Signals

Analysis of PCD signals measured from MBs is commonly performed in the frequency domain which exhibits characteristic emissions from microbubbles including the driving frequency, harmonics, subharmonics, ultraharmonics and broadband emissions. The information from the frequency spectrum helps to understand the cavitation phenomenon which is associated with the MB oscillation and disruption. The main characteristics of the frequency spectrum includes:

1. Fundamental or driven frequency (f)

At low stable cavitation regime when the MB is at rest or undergoing small oscillations, it emits the frequency spectrum of having frequency equal to that of driving frequency called the fundamental frequency. Usually this spectrum can be observed with or without MBs (water/tissue). As this signal is emitted from both tissue and MB, it is not useful on characterizing the MB behaviour.

2. Second harmonic (2f)

As the acoustic pressure amplitude increases, the MB oscillates nonlinearly emitting second harmonic (frequency spectrum having frequency twice that of driving frequency) of higher magnitude compared to that of tissue. This spectrum is associated with nonlinear propogation of ultrasound and has been exploited for harmonic imaging applications for the contrast enhancement (53). For therapeutic applications second harmonic emissions has to be explored for its usefulness.

3. Higher harmonics (nf)

At higher acoustic pressures, the MBs signals exhibit higher harmonics at 3f, 4f and so on for transmitted frequency f (4). The exact cause of these higher harmonics are not well understood, however it has been proposed that the nonlinear oscillation of the bubbles leading to chaotic behavior are associated with emission of harmonics (23).

4. Subharmonic (f/2) and ultraharmonics (mf/2)

Oscillating bubbles have potential to emit subharmonic peaks at integer fractions of driving frequency e.g f/2 and ultraharmonic peaks at (mf/2) where m is 3, 5.... for transmitted frequency f, which are not possible in tissue. These unique markers of the MB characteristics are associated with bubble dynamics. Investigations on sub-harmonics imaging (54; 44) and in therapy (45) have shown that subharmonic emission can be exploited as indicator of sonoporation. Although the mechanism behind subharmonic-ultraharmonic emissions are not well understood, four possible explanation have been suggested for free bubbles: (i) transient cavitation, (ii) subharmonic oscillation of bubbles excited at resonance, (iii) resonant oscillations of bubbles excited at their second harmonic and (iv) non-radial modes of oscillation. These phenomena of subharmonic and ultraharmonics emissions are described using bifurcation diagram (23) where bubble undergoes period doubling, period of four and eight. For better understanding detail investigation on sub-harmonics detection and its application has to be explored.

5. Broadband signal

Further increase of the pressure amplitude will cause the MBs to increase its size more than two times its initial radius and finally collapse or rupture. This phenomenon of bubble collapse occurs within a very short period of time where broadband spectra are emitted from MBs which are associated with post excitation signals (26; 55). In therapeutic applications this has been studied for inducing bioeffects (56; 57; 39). Figure 1.4 replicated from (58) represents the FFT response of MBs extract in frequency domain.

1.4.5 Cavitation Dose

Bioeffects induced by ultrasonically-stimulated MB have been associated with cavitation dose (57; 56; 39). Cavitation dose represents the amount of acoustic energy that is contained in the desired frequency window and is obtained by plotting the RMS FFT amplitude vs time (56). Furthermore the cumulative/integrated cavitation dose can be obtained from the integration of the entire RMS FFT amplitude vs time curve over the exposure time (decribe detail in section 2.4.2). In this project we estimate the cavitation dose and integrated cavitation dose using equations (2.2) and (2.3).



Figure 1.4: Figure: Showing the frequency spectra: fundamental (f), subharmonic (f/2) ultraharmonics (3f/2,5f/2) and second harmonic (2f) from MBs measured with acoustic method.

Numbers of studies were performed in vitro (39) and in vivo (59; 60) conditions. The inertial cavitation dose as a function of acoustic pressure, MBs concentration, and pulse length on hemolysis (vitro) and endothelial cell damage (vivo) have been studied. The conclusion was that both hemolysis and endothelial damage increases with cavitation dose. Similarly correlation of inertial cavitations as a function of applied pressure, bubble concentration and pulse duration with sonoporation have been investigated by (57; 56) where they have shown the cavitation dose correlates with pore sizes and sonoporation. All these experiments were performed with broad size distribution of MBs which strongly affects their emissions. Moreover, these experiments were performed under different exposure conditions and thus variation in cavitation dose and their resulting bio-effects was observed. Thus rigorous investigation on the MB acoustic behaviors controlling the exposure parameters and corresponding cavitation dose have to be optimized/controlled for producing useful bio-effects to improve sonoporation applications. Therefore the mo-

tivation behind the study is that the understanding of acoustic response of MB and their quantification at various acoustic pressure and pulse duration may potentially improve and monitor the ultrasound-MB mediated sonoporation applications.

1.5 Thesis outline

The hypothesis guiding this research is that the acoustic pressure and pulse duration have significant effect on MB acoustic behaviour and cavitation dose.

The main objective of this thesis is to characterize the acoustic behaviour of MBs at varying exposure conditions using the passive cavitation detection method.

The specific objectives of this study are:

- To measure the acoustic emission from MBs using passive cavitation detection at various pressures and pulse durations and study their response through FFT spectral analysis.
- To investigate the effect of ultrasound pressure and pulse duration on different MBs (Artenga and Definity) acoustic response.
- To quantify these bubble response through cavitation dose method.

The long term goal is to develop a monitoring tool of MBs activities during ultrasound treatment based on the PCD method, with aim to improve applications of drug /gene delivery using ultrasound.

Ultrasound and MB, their applications, MB response and their quantification and an outline of thesis is discussed in first chapter. In the second chapter, the PCD experimental setup and protocal, data acquisition and their analysis is discussed. Results are shown for MB behaviours and cavitation dose, as a function of acoustic pressure and pulse duration in third chapter. The fourth chapter contains a discussion, limitation of study and possible clinical relevance. The fifth chapter includes conclusions and directions for future work, with results for effect of MB sizes on their acoustic behaviours and appendix includes additional data to support results in chapter 3.

Chapter 2

Materials and Methods

PCD experiments were conducted with MBs (Artenga and Definity) in suspensions exposed to ultrasound at various acoustic pressure and pulse duration. The MBs response were analyzed in terms of frequency contents, and their quantification was performed through cavitation dose methods.

2.1 Experimental setup and procedures

2.1.1 Ultrasound Exposure Condition and Passive cavitation detection

A schematic diagram of the experimental setup for passive cavitation measurement is shown in Figure (2.1).

It consist of a programmable function generator (WW5062 Tabor Electronic Ltd, NY, USA) that transmits the short sequenced trigger controlled sine pulses (5 and 30 cycles) for insonation time of 1 s at a constant pulse repetition frequency (PRF) of 1KHz.



Figure 2.1: Schematic diagram of the PCD experimental setup. The bubbles emissions were received passively with 0.5 and 2.25 MHz transducers with an idea of detecting subharmonic and harmonics. The pulses were transmitted with 1 MHz transducer.

The pulses were transmitted to the RF power amplifier (240L, 50dB, ENI Inc. NY, Rochester, USA) that amplified the signals and drove a single-element spherically focused 1MHz transducer (IL0108HP, Valley Fisher Inc, Hopkinton, MA). PCD experiments were performed at room temperature in degassed water and with 3D micro-positioning systems graduated in mm scale for transducer positioning and 3D movable sample holder to ascribe its position. The sample was placed in a cylindrical tube at the focal volume which had dimensions of 10mm in length and 8mm in diameter. It was sealed with the mylair membrane and a magnetic stirrer was placed underneath for uniform mixing of the MBs suspension. The bubble emissions were collected passively by two PCD transducers at 2.25 MHz and 0.5 MHz (Valley Fisher Inc., Hopkinton, MA), that were positioned at 90 degrees to the treatment transducer at the focus as shown in the Figure 2.2. The PCD transducers were aligned perpendicular to the treatment transducer that enabled high spatial control over the region for which signals were collected because scattered signals only came from the MBs in the small overlapping confocal region. The acoustic emission signals collected from the MB suspension were amplified with a low noise (RF) amplifier (60dB, Miteq, USA) and passed through the attenuator (NAT-20, 20dB) and low pass filter (3.5 MHz) or high pass filter (1.25 MHz) to the computer.



Figure 2.2: A photograph showing the transducer alignment at the focus. The two receiver transducers on left and right were perpendicular to the source transducer. The sample holder was sealed with mylair membrane. All the devices had 3D micropositioning system graduated in mm scale for their positioning.

2.1.2 Exposure protocol

Before any experiments, a water tank of dimension $(50 \times 30 \times 20)$ cm³ was half filled with degassed water. The calibration for the focus and positioning of transducers was performed using the 3D positioning calibrated system and with standard technique of cross needle measurement. The maximum value of the pulse-echo signal from needle, seen on the oscilloscope when the ultrasound is on, determines the centre of the focus. The focus was at a distance of 4.9 cm, and 4.3 cm away from 2.25 and 0.5 MHz receiver transducers and 4.5 cm away from 1 MHz treatment transducer. The magnetic stirrer was placed underneath sample holder for the uniform mixing of the MB suspension. Water sample (1 ml) in the absence of MBs was placed with syringe, inside the sample holder tube which was constructed with mylair membrane sealed around it at the focus. The cylindrical shaped focal volume has a dimension of 10×8 mm. The power amplifier was turned on and computer program was initiated to record the acquired signals in acquisition card. The exposure was started by triggering the function generator with sequenced pulse containing PRP of 1 ms, repeated 1000 times for the total exposure time of 1 s (Figure 2.3). Experiments were conducted varying acoustic pressure (PNP) of 0, 0.5, 1.1, 1.5 MPa and pulse duration of $5\mu s$ and $30\mu s$. The transmit pressure amplitude was varied by controlling the function generator output voltage (0-600 mV) through Arb connection software (Tabor Electronic Ltd, NY, USA). After water signals (reference acquisition data) recording were completed the MB samples were prepared at 1% concentration and were placed in the focal volume and the signals were recorded for the given exposure conditions. For each of the exposure conditions the sample was replaced with new MBs sample and the signal received with transducers were acquired in data acquisition system.



Figure 2.3: Diagram showing the exposure pulse sequence and data acquisition timing. PD : pulse duration (5 and 30μ s), PRP : pulse repetition period (1ms), TD : trigger delay (60μ s) and AW : acquisition window (48μ s)

2.1.3 Data Acquisition

The acoustic signals emitted by MBs and water were recorded in 1 ms time intervals, after the sequenced pulse was initiated (Figure 2.3). A total of 1000 signals (each having 6000 samples) were acquired at a sampling rate of 125 MHz in an Alazar DSO acquisition system (ATS460-128M-S600726) with an acquisition window of length 48 μ s for given exposure time of 1 second. Each acquisition, $s_i(t)$ where i = 1 to 1000 represents the acquisition events for each time points. The time delay of 60 μ s was applied with respect to the trigger signal, which was necessary to account for the travel time from the source to the receiver transducers. The acquisition was stopped after acquiring the data for 1 s and the data was saved as MATLAB file for further analysis.

2.1.4 Transducer Specifications

Three transducers were used in this project. A spherically focused single element 1 MHz transducer (IL0108HP, Valpey Fisher Inc, Hopkinton, MA) with a diameter of 25 mm, focal length of 4.5 cm and bandwidth of ± 6 dB was used for transmitting ultrasound pulse to the MB suspension while a 0.5 MHz (IL0508HP, Valpey Fisher Inc, Hopkinton, MA) and a 2.25 MHz (IL0208HP, Valpey Fisher Inc, Hopkinton, MA) transducers were used for measuring the acoustic signals from MBs. Both have a diameter of 25 mm, focal lengths of 4.3 and 4.9 cm respectively and the bandwidth of ± 6 dB.

2.1.5 Transducer calibration

All transducers were characterised using a calibrated needle hydrophone (Precision Acoustics Ltd, Dorchester, and Dorset, UK). In absence of an exposure chamber the pressure was measured at the focus of the transducer beam. The pressure field was characterized based on the graph shown below for 1 MHz transducer (Figure.2.4). The peak negative pressure was used to characterize the acoustic amplitude.

2.2 Microbubble agent

Two MB agents (Definity and Artenga) were used in this study. Definity, Perflutren lipid microspheres, (Lantheus Medical Imaging, Billerica, MA, USA) are FDA approved ultrasound imaging contrast agents composed of the octofluoropropane (C3F8) gas core stabilised by phospolipid shells. The container vials have a concentration of 1.2×10^{10} bubbles/mL, and average diameters of 1.1-3.3 μ m, with 98% less than 10 μ m according to the manufacturer. The Definity vial was activated by shaking it with Vial-Mix (Lan-


Figure 2.4: 1 MHz transducer calibration curve. Y- axis representing pressure in MPa. The X-axis represents voltage in mV. The Green curve is peak-to-peak pressure, the red curve absolute value of peak negative pressure and the blue curve peak positive pressure.

theus Medical Imaging, Billerica, MA, USA) for 45 seconds and was equilibrated for 5 minutes at room temperature. The vial was agitated for 10 seconds by hand, inverted for 30 seconds and was vented with an 18-gauge needle. Finally MBs were transferred to a syringe and were diluted in water at the desired concentration. Artenga MB, an ultrasound contrast agent, currently in clinical trials contains perflurocarbon gas enclosed in a lipid shell. It has a mean bubble diameter of 2 microns and a concentration of 1×10^9 microbubbles/mL (Artenga Inc, Ottawa, canada). The MB suspension was prepared according to the manufacturer instructions (Artenga Inc, 2008).

A Coulter Counter System (Multisizer IV, Beckman Coulter Inc, Fullerton, CA) was used for the MB size distribution measurements. The size distribution of MBs in a 5 μ L volume were measured using a 20 μ m aperture tube. Measurements were performed for Artenga MB having volume weighted diameter of 1-20 μ m in range. Figure 2.5 shows the Coulter Counter results where the average MBs size distribution was approximately 2-4 μ m.



Figure 2.5: Volume weighted size distribution measurements with Coulter Counter for Artenga MBs .

2.3 Experiments performed

PCD experiments were performed with two MBs (Artenga and Definity) at various acoustic pressure, pulse duration and microbubble sizes. The parameters used in the experiments are discussed in previous subsection (2.1.1). The list of PCD experiments performed during study period of 2 years with some specifications are shown in Table 2.1

Table 2.1: List of PCD experiments: where PD=pulse duration, PNP= Peak negative pressure and FIL/UNF represents filtered/unfiltered bubbles and NOE represents numbers of experiments.

Exp.No	MB type	PD(us)	PNP(MPa)	Receiver(MHz)	FIL/UNF	NOE
1	ART	30	0-1.5	2.25 and 0.5	UNF	3
2	DEF	30	0-1.5	2.25 and 0.5	UNF	3
3	ART	5	0-1.5	2.25 and 0.5	UNF	3
4	DEF	5	0-1.5	2.25 and 0.5	UNF	3
5	DEF	30	0-1.5	2.25 and 0.5	FIL	1

2.4 Data Analysis

2.4.1 Microbubble behaviour

The acquired acoustic signals ($s_i(t)$ from MBs at 1ms time interval were analyzed using MATLAB (Mathworks, Natick, MA). The post triggered samples (24-48us) were analyzed using the rectangular window for the frequency response. The frequency response were obtained with FFT algorithm in MATLAB using the transformation equation $[S_i(f)] = 20 \log_{10} \text{ FFT}[s_i(t)]$. At 10 ms after exposure the equation used was

$$[S_{10}(f)] = 20 \log_{10} \text{FFT}[(s_{10}(t)]$$
(2.1)

Moreover the MB acoustic response was analysed by plotting spectrograms which showed the time dynamics of MB emissions and their activities in term of frequency content over insonation time. Furthermore cavitation MB activities were measured in terms of cavitation dose.

2.4.2 Cavitation dose

The response of MBs varied with insonation time. The cavitation dose of MB response was calculated based on the RMS FFT Algorithm as described by (39). The mathematical expression for calculating cavitation dose in this project is given as

$$CD_{RMS} = \int_{f-\Delta f}^{f+\Delta f} |S_i(f)|^2 df$$
(2.2)

where i=1-1000 represents the acquisition events for each time points, $S_i(f)$ represents the FFT of $s_i(t)$. $\Delta f = 0.2$ MHz and f = (0.5, 1.5, 2 MHz for the subharmonic, ultraharmonic and second harmonic frequency spectrum respectively). The interval between f- Δf and f+ Δf represents the frequency window for calculating the cavitation dose.

In this project the RMS FFT amplitude vs time plot (cavitation dose) was calculated for 1000 signals each acquired at 1 ms time interval at desired frequency windows of subharmonic $(0.5\pm0.2\text{MHz})$ second harmonic $(2\pm0.2\text{MHz})$ and ultraharmonic $(1.5\pm0.2\text{MHz})$ frequencies. In addition the integrated cavitation dose (ICD) at given frequency windows over the exposure time of 1 s were obtained following an idea as discussed by(56; 39). The ICD for each spectrum (subharmonic, harmonic, and ultra-harmonics response) was defined as the area under the RMS FFT amplitude curve over the exposure time of 1 s. The following mathematical expression was used to calculate the integrated cavitation dose

$$ICD = \sum_{i=1}^{1000} |CD_{RMS}| \Delta t \tag{2.3}$$

where Δt represents the time difference between two acquisition events which is 1 ms. A schematic diagram representing data analysis algorithm is shown in Figure 2.6



Figure 2.6: Data analysis algorithm: (a) Time domain signal at 10 ms $[s_{10}(t)]$. (b) represents frequency response of TD signal obtained using equation 2.1. (c) showing the frequency window at ultraharmonics and second harmonic frequency spectra for calculating the cavitation dose. (d) graph representing cavitation dose obtained using equation 2.1. (e) ICD was obtained integrating the cavitation dose from the shaded region over the exposure time of 1 s using equation 2.2.

Chapter 3

Results

In this chapter, the acoustic characteristics of the two MBs (DEF and ART) exposed at varying acoustic pressures (0-1.5 MPa) and pulse duration (5 and 30 cycles) for insonation time of 1s was investigated. The MB acoustic response are shown in Section 3.1 and the cavitation dose results in Section 3.2.

3.1 MB acoustic behaviour

3.1.1 Effect of acoustic pressure (PNP)

The effect of acoustic pressure (PNP) on acoustic MB behaviours are shown in Figures 3.1 to 3.4 which presents the PCD results for different PNP. MBs exhibit nonlinear and chaotic behaviours (Figure 3.1 and 3.2). Spectrograms associated with the acoustic response of the MB are presented in Figure 3.3 and FFT spectra at various time points are shown in Figure 3.4.

The acoustic behaviour of ART MB exposed to 30 cycle pulses at PNP (0-1.5 MPa) depends on the applied PNP (Figure 3.1 and 3.2). The time domain signals received



Figure 3.1: ART MB behaviours (red) and water (blue) as a function of PNP at 10 ms after exposure. Figures (a,c,e,g) and (b,d,f,h) represent the time domain signals and their corresponding FFT response at (0,0.5,1.1,1.5) MPa respectively. The vertical axis and horizontal axis for TD signal represents amplitude (arbitrary units) and time (s) while it is pressure amplitude (dB) and frequency (Hz), for FFT figures. MBs and water were insonated with 30 cycle 1 MHz pulse at PRF of 1 KHz and were received with 2.25 MHz transducers with highpass filter of 1.25 MHz.

with the 2.25 MHz transducer and their corresponding FFT spectra are shown in Figure 3.1 at10 ms after MB were exposured to ultrasound. At 0 MPa, the MB exhibits no sign of oscillation and its corresponding FFT spectrum is featureless Figure 3.1(a,b). At 0.5 MPa, the MB shows weak non-linear oscillation with an emission of the fundamental (f), second harmonic (2f) and third harmonic (3f) Figure 3.1(c, d). The pressure amplitude are higher than water by 67 dB for (2f) and 45 dB for (3f). At 1.1 MPa, the MBs undergo strong nonlinear oscillation with an emission of second harmonic (2f), ultra-

harmonics (1.5f, 2.5f) and third harmonic (3f) greater than water by 73, 76, 91, and 52 dB respectively Figure 3.1 (e, f). At 1.5 MPa, the MBs exhibit chaotic behavior with an emission of broadband spectra containing second harmonic (2f), ultra-harmonics (1.5f, 2.5f) and third harmonics (3f) by 98, 83, 66 and 67 dB greater than water respectively (Figure 3.1 (g, h)). More detail analysis of the time domain signal is discussed in the appendix (Figure A.1).



Figure 3.2: ART MB behaviours (red) and water (blue) as a function of PNP at 10 ms after their exposure. Figures (a,c,e) and (b,d,f) represent the time domain signals and their corresponding FFT spectra at (0.5, 1.1, 1.5) MPa respectively. The vertical axis and horizontal axis for TD signal represents amplitude (arbitrary unit) and time (s) while it is pressure amplitude (dB) and frequency (Hz), for FFT figures. MBs and water were insonated with 30 cycle 1 MHz pulse at PRF of 1 KHz and were received with 0.5 MHz transducers with lowpass filter of 3.5 MHz.

The time domain signals received with the 0.5 MHz transducer for 10 ms insonation

time after exposure of MB to ultrasound and their corresponding FFT spectra are shown in Figure 3.2. At 0.5 MPa, the MBs exhibit weak non-linear oscillation (unequal and irregular expansion and contraction) with an emission of the subharmonic (f/2) of magnitudes 21 dB higher than water Figure 3.2 (a, b). At 1.1 MPa, the MBs undergo asymmetric oscillations which exhibit expansion-like behaviour and emit the subharmonic frequency spectrum of magnitude 18 dB greater than water Figure 3.2 (c, d). At 1.5 MPa, the MB exhibits compression-like behavior and emit the subharmonic frequency spectrum of magnitude 30 dB higher than water Figure 3.2 (e, f). More detail analysis of the time domain signal is discussed in the appendix (Figure A.2).

The time dynamics of ART MB acoustic behaviour, received with 0.5 MHz and 2.25 MHz transducers, are presented in spectrograms Figure 3.3. It is seen that with an increase of pressure, nonlinear behaviour of the MBs increases. The time dynamic results indicate that MBs undergo large scale oscillations with emission of the higher amplitude frequency spectra up to 200 ms following the lower amplitude signals with longer insonation times Fig 3.3 (b, d, f). At 1.1 MPa, the frequency spectra from the MBs nonlinear oscillation with respect to insonation time are observed to be very strong during the first 200 ms. At 1.5 MPa, the MBs exhibit spectra of higher magnitude for the first 100 ms. Following this, the signals disappear for 200 ms and reappear at 300 to 500 ms Figure 3.3 (f) as shown by the arrow. The spectrograms for water at 1.1 MPa Figure 3.3 (h) show no other frequency spectrum except the fundamental frequency (f). Similar results from experiment 2 and experiment 3 are shown in appendix (Figure 5.8). Spectrograms with the 0.5 MHz transducer have shown that the MBs frequency spectra for subharmonic have greater amplitude as the pressure increases. These results are from different experiments than that received with 2.25 MHz transducers.

The frequency spectra from ART MB, detected with the 2.25 MHz transducer at



Figure 3.3: ART MB spectrograms as a function of PNP for signals received with 0.5 and 2.25 MHz receiver transducers. Results are shown for (0.5, 1.1, 1.5) MPa for MB Figure 3.3 (a to f), and water at 1.1 MPa (g,h). The vertical axis on the spectrogram represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.

different time points (Figure 3.4) demonstrate that the FFT peak pressure amplitude increases with the increase of pressure while it decreases with insonation time. The time points were chosen based on the result of the spectrogram Figure 3.3. At 0.5 MPa, the peak pressure amplitude of the second (2f) and third harmonic (3f) decreases from 101 to 55 dB, and 70 to 43 dB respectively, within 240 ms insonation time. At 1.1 MPa, it decreased from 103 to 15, 110 to 74, 101 to 13 and 98 to 38 dB within an insonation time of 300 ms, for the ultraharmonic (1.5f), second harmonic (2f), ultraharmonic (2.5f) and third harmonic (3f) frequencies respectively. At 1.5 MPa, the peak pressure amplitude of



Figure 3.4: FFT spectra of ART MB at various time points for the data received with 2.25 MHz transducers. The signals in blue colour represents FFT spectra at various time points for 0.5 MPa, red for 1.1 MPa and green for 1.5 MPa. The vertical axis represents pressure amplitude (dB) and horizontal axis represents frequency (Hz).

the ultraharmonic (1.5f), second harmonic (2f), ultraharmonic (2.5f) and third harmonic (3f) decreased from 116 to 50,127 to 65, 108 to 45, 95 to 32 dB respectively within 500 ms insonation time.



3.1.2 Effect of acoustic pressure and pulse duration on MB type

Figure 3.5: FFT spectra for ART MB at 10 ms insoonation time and DEF MB at 43 ms insonation time (for30 cycles) and 210 ms insonation time (for5 cycles) after exposure and PNP of 0.5, 1.1, 1.5 MPa. Blue color represent 5 cycles and red colour represents 30 cycles pulse duration, and the signals were received with 2.25 MHz transducer. The vertical axis represents pressure amplitude (dB) and the horizontal axis represents frequency (Hz).

The frequency spectra of both MBs (Artenga and Definity) exposed to 30 cycle and 5 cycle pulse duration at acoustic pressure of 0.5, 1.1 1.5 MPa are shown in Figure 3.5. ART MB FFT spectra for both pulse duration are shown at 10 ms insonation time after exposure while DEF MB FFT spectra are shown at 43 ms for 30 cycle pulse duration and 210 ms for 5 cycle pulse duration after exposure. These time points for DEF MB were chosen based on the results shown in Figure 3.7 where the MB activities were significant to analyse and 10 ms was choosen based on Figure 3.3 and 3.6. Results demonstrate that the FFT spectrum of 5 cycle exposure were different from 30 cycle exposure. Similar results from second set of experiment are shown in appendix (Figure B.6).

The ART and DEF MBs, both at 5 cycles pulse duration have broader FFT spectra with lower pressure amplitudes compared to 30 cycles frequency responses Figure 3.5. Large difference in the pressure amplitude can be seen with a range of 8 to 47 dB among the harmonics and ultraharmonics. At 5 cycle pulse duration ultraharmonic signal (1.5 MHz) of lower amplitude was seen at 0.5 MPa (Figures 3.5 and B.6). This can be clearly seen from the spectrograms results (Figures 3.7, B.3 and B.4) where lower amplitude frequency spectrum is visible at 1.5 MHz. For 30 cycle pulse duration results, this signal was neither seen in FFT spectra (Figures 3.5 and B.6) nor in spectrograms (Figures 3.3, B.1 and B.2).

In addition ART and DEF MBs at 5 and 30 cycles pulse duration have different frequency response (Figure 3.5). The frequency spectra for Definity MB (5 cycle) have the higher peak pressure amplitude of ultraharmonic (1.5f), second harmonic (2f), ultraharmonic (2.5f) and third harmonic (3f) than the corresponding Artenga MBs spectra at 0.5 and 1.1 MPa. The difference in the peak pressure amplitude was 17, 11, and 16 dB at 0.5 MPa (no 2.5f) and 3, 4, 5, and 1 dB at 1.1 MPa. However at 1.5 MPa it was opposite and the difference in peak pressure amplitude were 3, 20, 18, and 39 dB higher for ART MBs.

For 30 cycle pulse duration, ART MBs have higher peak pressure amplitudes than Definity for most of the FFT spectra at all pressures (Figure 3.5). The peak pressure amplitude differences were 11 and 8 dB for second (2f) and third harmonics (3f) at 0.5 MPa. At 1.1 MPa the differences were 7, 1, 11, and 2 dB for the ultraharmonic (1.5—emphf), second harmonic (2f), ultraharmonic (2.5f) and third harmonic (3f) respectively,



Figure 3.6: Spectrograms for ART MB, at 5 cycles pulse duration and PNP (0.5, 1.1, 1.5 MPa) for the signals received with 2.25 MHz transducer. The vertical axis represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.

while they are 13, 25, 3 and 12 dB for the ultraharmonic (1.5f), second harmonic (2f), ultraharmonic (2.5f) and third harmonic (3f) respectively at 1.5 MPa. In addition the spectrograms comparison between DEF and ART at 30 cycle (Figures 3.3, 3.7, B.1, B.2) have shown the frequency amplitude are greater for ART which support our results that ART have greater peak pressure amplitudes than DEF in FFT plots (Figure 3.5 and B.6)

The spectrogram (Figure 3.6) shows the time dynamic behaviour of the ART MBs for five cycles exposure, which were received with 2.25 MHz transducer. It exhibits a wide range of frequency spectra distribution for first 200 ms of insonation time at 1.5 and 1.1 Mpa. At 0.5 Mpa, the frequency contents are of low magnitude and greatly varies with time. As insonation time increases, the spectra (f, 2f, 3f) have very low frequency amplitude compared to that of 30 cycles results shown in Figure 3.3 (b, d, e). Similar results from experiment 2 and experiments 3 are shown in appendix (Figure B.3).



Figure 3.7: Spectrograms for DEF MB, at 5 and 30 cycles pulse duration and PNP (0.5, 1.1, 1.5) MPa for the data received with 2.25 MHz transducer. The vertical axis represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.

The time dynamics of frequency spectra for the DEF MB at 5 cycles and 30 cycles pulse duration are shown in Figure 3.7. The spectrograms demonstrate that the signal from the MBs when exposed to 5 cycles, have variable frequency amplitude for total exposure time of one second, with signals of higher magnitude at around 200 ms while the signals for 30 cycles pulse duration are observed with higher magnitude at 45 to 80 ms and there are no signals after 200 ms except f and 2f of small magnitude. Similar results from experiment 2 and experiment 3 are shown in appendix (Figure B.2 and B.4).

3.2 Cavitation Dose

The cavitation dose plots (RMS FFT amplitude vs time) for ART MB at 30 cycles insonation pulse and PNP of 0.5, 1.1 and 1.5 MPa are shown in Figure 3.8. Cavitation doses are calculated at the desired frequency windows $(0.5\pm0.2, 2\pm0.2, 1.5\pm0.2, 2.5\pm0.2)$ MHz) as the subharmonic cavitation dose, second harmonic cavitation dose and ultraharmonics cavitation dose respectively. Results demonstrate that the cavitation dose increases as a function of pressure. At 1.5 MPa, the subharmonic (f/2) cavitation dose is greater (40 units) at 5 ms and decreases significantly to 2 units at 100 ms, following the lower values with increase of insonation time (Figure 3.8 (a)). The ultraharmonic (1.5f)cavitation dose has shown that it grows with insonation time, attains a maximum value of 43 units at 34 ms and decays slowly to 2 units at 120 ms (Figure 3.8 (b)). Further, the second harmonic cavitation dose plots have indicated that it grows to 74 units at 30 ms and decreases to 5 units at 200 ms (Figure 3.8 (c)). The ultraharmonic cavitation dose plot demonstrate that the cavitation dose suddenly grows higher at 10 ms, maintains its amplitude up to 50 ms (average of 17 units) and finally decreases to 2 units at 120 ms. Similar decrease of cavitatitation dose with increase of insonation time can be seen in Figure 3.8 at 1.1 and 0.5 MPa. The cavitation dose follows the similar trend of increase with PNP and decrease with insonation time for all others experimental results. Some of the results are shown in appendix (Figure B.7 and B.8). The water signals (blue in colour) at 1.1 MPa at each frequency window were shown as a reference signal.

Integrated cavitation doses for DEF and ART MBs at 30 cycle pulse duration are shown in Figures 3.9 and 3.10. The error bar represents the mean \pm standard deviation for signals from 3 experiments. The ICD increases with increase of PNP for each spectral feature. The subharmonic ICD is saturated for DEF MB at 1.1 MPa while large growth



Figure 3.8: RMS FFT amplitude vs time plots for ART MB at 30 cycles pulse duration. (a), (b),(c) and (d) represent the cavitation dose for subharmonic $(0.5\pm0.2 \text{ MHz})$, ultraharmonic $(1.5\pm0.2 \text{ MHz})$, harmonic $(2\pm0.2 \text{ MHz})$ and ultraharmonic $(2.5\pm0.2 \text{ MHz})$ respectively. The vertical axis represents the RMS FFT amplitude (arbitrary unit) and horizontal axis represents the time (s).

of subharmonic at 1.1 MPa can be seen for ART MB. In addition the ICD values at each pressures for subharmonics are greater for DEF MBs (Figure 3.9). The cavitation dose for ultraharmonic and second harmonic are greater for DEF MBs. The cavitation dose for DEF MBs are saturated at 1.1 MPa while strong growth of ultraharmonic cavitation dose for ART MBs is observed (Figure 3.10).

Integrated cavitation doses for DEF and ART MBs at 5 cycle pulse duration are shown in Figure 3.11 and Figure 3.12. The error bar represents the mean \pm standard deviation for signals from 3 experiments. The ICD increases with increase of PNP for



Figure 3.9: Integrated cavitation dose for DEF and ART MBs for subharmonic $(0.5\pm0.2 \text{ MHz})$ frequency spectrum for signals received with 0.5 MHz transducer, at acoustic pressure of 0.5, 1.1 and 1.5 MPa and 30 cycle pulse duration. The vertical axis represents the ICD (arbitrary unit) and horizontal axis represents subharmonic frequency spectra at different pressures. The error bar represents the mean± standard deviation for signals from 3 experimentals.

each spectral feature. The subharmonic ICD is close to saturation level for DEF MB at 1.1 MPa while large growth of subharmonic ICD is seen for ART MB between 1.1 and 1.5 MPa. In addition the ICD values at all pressures are greater for DEF MBs except at 1.5 MPa (Figure 3.11). The cavitation dose for ultraharmonic and second harmonic are greater for DEF MBs. The harmonic cavitation dose for DEF MBs is saturated at 1.1 MPa . The small growth of ultraharmonic cavitation dose for ART and DEF MBs is observed at 1.1 and 1.5 MPa (Figure 3.12).

Results demonstrate that the integrated cavitation dose values are greater for DEF MB for most of the frequency components than corresponding frequency spectra for ART MB. This implies that the higher cavitation dose of DEF at 5 cycle pulse duration and



Figure 3.10: Integrated cavitation dose for DEF and ART MBs for ultraharmonic $(1.5\pm0.2 \text{ MHz})$ and second harmonic $(2\pm0.2 \text{ MHz})$ frequency spectra for signals received with 2.25 MHz transducer, at acoustic pressure of 0.5, 1.1 and 1.5 MPa and 30 cycle pulse duration. The vertical axis represents the ICD (arbitrary unit) and horizontal axis represents frequency spectra at different pressures. The error bar represents the mean \pm standard deviation for 3 experiments.

higher pressure may be useful for inducing bio-effects while higher and more pronounced acoustic response from ART MBs at 30 cycle pulse duration may be a good choice for MB activities monitoring purpose.



Figure 3.11: Integrated cavitation dose for DEF and ART MBs for subharmonic $(0.5\pm0.2 \text{ MHz})$ frequency spectrum for signal received with 0.5 MHz transducer, at acoustic pressure of 0.5, 1.1 and 1.5 MPa and 5 cycle pulse duration. The vertical axis represents the ICD (arbitrary unit) and horizontal axis represents frequency spectra at different pressures. The error bar represents the mean \pm standard deviation for 3 experiments.



Figure 3.12: Integrated cavitation dose for DEF and ART MBs for ultraharmonic $(1.5\pm0.2 \text{ MHz})$ and second harmonic $(2\pm0.2 \text{ MHz})$ frequency spectra for signals received with 2.25 MHz transducer, at acoustic pressure of 0.5, 1.1 and 1.5 MPa and 5 cycle pulse duration. The vertical axis represents the ICD (arbitrary unit) and horizontal axis represents frequency spectra at different pressures. The error bar represents the mean \pm standard deviation for 3 experiments.

Chapter 4

Discussion

The MB acoustic behaviours of DEF and ART were studied for various acoustic pressures and pulse durations using passive cavitation detection measurements. The time domain signals and their corresponding FFT spectra were analysed to characterise their acoustic response. Spectrogram analysis was employed to investigate their time dynamic response to ultrasound and a cavitation dose technique was used for their quantification.

4.1 Nonlinear Oscillation

Results have demonstrated that the nonlinear response of the MBs increases with the PNP. At 0 Mpa, the MB exhibited no sigh of oscillation (Figure 3.1) which demonstrated that certain pressure threshold is requided for its oscillation. The non-linear oscillation regime of the MB was identified for the acoustic pressure between 0.5 to 1.5 MPa. It was shown that the asymmetric, irregular and aperiodic oscillations of the MB increased with PNP, that emitted frequency spectra at the subharmonic, ultraharmonics, harmonics, and broadband spectra (Figures 3.1 and 3.2). Moreover, only above certain

pressure thresholds, the characteristic markers of the nonlinear oscillation of MB such as sub-harmonics and ultra-harmonics are detected. These MBs characteristics frequency spectra (subharmonics and ultraharmonics) are believed to be associated with the bubble motion undergoing period doubling, period of four and period of eight as shown by using the bifurcation diagram analysis (23).

We have shown that with pulse duration, the acoustic behaviour of MBs greatly vary. Five cycle pulse duration results had very broad and lower peak pressure amplitude spectra by 8-47 dB than 30 cycles pulse duration. This is likely due to reason that the wide range of MBs are oscillating emitting the broad frequency spectrum from a broadband pulse (5 cycle). The narrowband pulse (30 cycle) is more specific to certain sizes of MBs, due to its small bandwidth. In addition, the damping and attenuation is greater for 5 cycle pulse as more MBs are responding to ultrasound pulse which cause the peak pressure amplitude are lower for 5 cycle pulse than that of 30 cycle pulse. It was observed that the ultraharmonic frequency spectrum (1.5f) appear at 0.5 MPa for the 5 cycle pulse duration (Figures 3.5 and B.6) and spectrograms (Figures 3.6, 3.7, B.1, B.2, B.3 and B.4) while there are no such signal for 30 cycle pulse duration. Moreover, the FFT response (Figures 3.5 and B.6) agree with the results shown by (45) where the effect of pulse duration on emissions of sub-harmonic spectra was investigated and has shown that higher spectral emission (subharmonic) was achieved for higher pulse duration. These results have demonstrated that the choice of 30 cycle pulse duration is better for the monitoring purpose, as the FFT signals for 30 cycles are very high and clear from noise level.

The effect of acoustic pressure and pulse duration on MB acoustic response has shown the FFT spectra for ART are wider while spectra for DEF are narrower. This could possibly happen because of difference in encapsulation (elasticity and viscosity), sizes and gas contents which may results different scattered strength.

Spectrogram results have shown the time dynamics of the MB frequency spectra. The strong MB non linear activities at 30 cycle pulse duration are detected for 200 ms and no signals except fundamental and second harmonic are seen for longer insonation time (Figure 3.3, 3.7, B.1 and B.2). The subsequent decrease in spectral signals after 200 ms is believed to indicate the loss of bubbles that are destroyed or loss of stability of MB due to diffusion of the gas, which reduce both scattering and emissions from MBs. Thus, fewer bubbles may not be efficient enough to show the significant bubble response. This provides us an idea about the life span of MB during insonation.

Spectrograms (Figures 3.6 and 3.7) demonstrate that the pulse duration effects the MB behaviours, i.e the broadband frequency spectra showing the MBs disruption phenomena at some time points were distinctly observed for 5 cycle pulse duration compared to 30 cycles pulse duration (Figures 3.6, 3.7, B.3 and B.4). In addition the MBs activities persisted for longer insonation time for 5 cycle pulse duration, which is the result of MBs oscillating continuously for longer period of time before get disrupted.

We have shown that there is significant difference in the strength of MB frequency spectral emissions and their time dynamics for the DEF (FDA approved) and ART (clinical trial) agents (Figures 3.6 and 3.7). The frequency spectra amplitude was higher for ART MB for first 200 ms while they delayed by 70 to 200 ms in case of DEF MBs. In addition, it is shown that DEF oscillates for longer time and get dusrupted, while ART have shown oscillation at the beginning of insonation time points and persists for almost 200ms for 5 cycle pulse duration. This could possibly happen because of difference in encapsulation (elasticity and viscosity), sizes and gas contents which may results different scattered strength and at different time points. Similar information can be seen in appendix Figures B.3 and B.4.

4.2 Cavitation dose

The cavitation dose (Figures 3.8, B.7, B.8, B.9, B.10) and integrated cavitation dose (Figures 3.9, 3.10, 3.11, 3.12) for each of the frequency content (subharmonic, second harmonic and ultraharmonics) increases with pressure which agrees with the published results shown by (57; 56; 39) where the integrated cavitation dose in the certain frequency window for broadband emissions were investigated and have shown that it increases with the pressures. As the pressure increases the MBs oscillates nonlinearly in large scale and at same time large numbers of the MBs get disrupted emitting the sound which give rise to higher integrated cavitation dose. The integrated cavitation dose was found saturated for second harmonic at 1.1 MPa which is due to the energy redistribution to the ultraharmonics spectra.

The integrated cavitation dose was obtained greater for 5 cycles pulse duration which contradicts the results of (60) where they have shown that with increase of pulse length integrated cavitation dose was higher. This could have occurred because it has been shown that the integrated cavitation dose for broadband emissions associated with the MBs destruction phenomenons was investigated by (60) while in our experiments the investigation was on the non-linear regimes where most of the bubbles undergoes nonlinear oscillating rather than destruction. The reason of having higher integrated cavitation dose at 5 cycles is that the spectra are broader at 5 cycles which content more acoustic energy, which was emitted from wide range of MBs. The longer MB activities seen in spectrogram at 5 cycles pulse duration support the results of having higher integrated cavitation dose. The cavitation dose was greater for DEF MBs than that of ART which possibly be the concentration and size effects which give more MB activities as seen for longer time as demonstrated in Figures (3.6, 3.7, B.3, B.4). This suggest higher number density of Definity with smaller sizes may oscillates at large scale compared to Artenga MB and thus give higher integrated cavitation dose. Therefore, Definity might be a right choice for inducing bioeffects because it has been shown that higher the cavitation dose more is the induced bioeffects.

Finally this work suggests two different future directions: In order to develop frequency spectra as monitoring tools, we require high amplitude acoustic spectra, that was achieved at intermediate (1.1 MPa) acoustic pressure and higher (30 cycle) pulse duration and for ART MBs while the higher integrated cavitation dose might correlates to enhance permeability which can improve sonoporation applications that was obtained at higher pressure but lower pulse duration and was greater for DEF MBs.

4.3 Limitations of this study

The MB preperation and conducting experiments for long hours was challanging. Using MBs suspension at time of generation and after half an hour make a difference of few dB in emissions spectrum which may influence the results of peak amplitude of frequency spectra. PCD experiments were performed throughout the period of two years and ecah time the MB viais used in the experiments were different. Since the MB vial kept for longer time from date of manufacture have shown weaker response compared to the fresh MBs, this might have influence on our results. Moreover the PCD experiments were conducted in water at room temperature which may not represent the complexity of the vivo tissue models. In addition, the water was used as a reference which cannot account for nonlinear propogation effects on MBs response. The bandwidth of the receiver trnsducers was not taken into account during the analysis of PCD data.

4.4 Possible applications and clinical relevance

Improving our nderstanding of MB behaviors and correlating them with induced bioeffects may enable us to increase efficacy of ultrasound and MB mediated applications. This project has potentially open a way of enhancing sonoporation activities and monitoring it based on the MB acoustic characterization. The acoustic methods developed in bench top system can be translated and tested in vivo models.

Chapter 5

Conclusion and Future work

5.1 Conclusion

The non-linear oscillation of MBs increases with acoustic pressure and pulse duration. MBs exhibits strong subharmonics, and ultraharmonics at 1.1 MPa at 30 cycle pulse duration while and at 0.5 MPa for 5 cycle pulse duration. MBs activities (oscillation and disruption) appear significant up to 200 ms for 30 cycle pulse duration, and appear longer for 5 cycle pulse duration. The integrated cavitation dose increased with acoustic pressure for all frequency components including subharmonic, ultraharmonics and second harmonic, and decreases with pulse duration. The ICD was greater for DEF MBs compared to ART MBs.

5.2 Future work: Effect of Microbubbles Size

In this section, the acoustic characteristics of the filtered DEF MBs exposed at varying acoustic pressure (0.5-1.5 MPa), pulse duration 30 μ s and different MB sizes for insonation time of 1 second was investigated under the same exposure conditions of PCD experiments that was described in chapter two. Methods includes the MBs size isolation through centrifuge which was performed based on the migration of MB in the centrifugal field(61). The MB suspension was centrifuged and MBs were acquired from different depths of a tube corresponding to small and large bubbles. Large MBs were at the top while smaller ones were at the bottom of the tube. Analysis of data was performed for ectracting spectrogram and cavitation dose methods. Spectrogram results are shown in Figures 5.1 and 5.2 and integrated cavitation dose in figure 5.3

5.2.1 Results and discussion

The acoustic behaviours of filtered DEF MBs exposed to 30 cycles pulse at acoustic pressure of 0.5, 1.1, 1.5 MPa and measured with 2.25 MHz transducer are shown in spectrograms Figure 5.1. The MBs exhibit different acoustic responses depending on size. Results demonstrate that the smaller size MBs exhibit higher frequency amplitude than the larger sizes. The time dynamic response is variable (appears and disappears) for filtered MBs within insonation time of 1 s while it is less than 100 ms for unfiltered MB and after that no signals are seen except fundamental and second harmonic (Figure 5.1).

The signals are received with 0.5 MHz transducer, where we are interested at subharmonic frequency response. At 0.5 Mpa, the frequency amplitude for subharmonic spectrum was lower for large MBs compared to smaller one while at 1.1 MPa it was greater than for larger MBs. At 1.5 MPa the frequency amplitude for subharmonic was greater for the smaller MBs for 200 ms and slowly decays.

The ICD are plotted for the subharmonic $(0.5\pm0.2 \text{ MHz})$, ultraharmonic $(1.5\pm0.2 \text{ MHz})$



Figure 5.1: Spectrograms for filtered DEF MBs, at 30 cycle pulse duration and PNP (0.5, 1.1, 1.5 MPa) for the signals received with 2.25 MHz transducer. The vertical axis represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.

MHz), and second harmonics $(2\pm0.2 \text{ MHz})$ as a function of acoustic pressure and MB sizes (Figure 5.5). It was observed that the cavitation dose for all frequency components increases with acoustic pressure. Large scale subharmonic, ultraharmonic and second harmonic growth is observed at 1.1 and 1.5 MPa for large, small and unfiltered MBs. The ICD growth is relatively greater for larger MBs at 1.1 MPa while it is greater for small MB for all others exposure pressures. Moreover the ICD for second harmonics are greater than subharmonics followed by ultraharmonics.

Spectrograms have shown that the time dynamics greatly varies for different bubbles sizes. Smaller the MBs higher are the spectral amplitude and persisted for longer in-



Figure 5.2: Spectrograms for filtered DEF MB, at 30 cycle pulse duration and PNP (0.5, 1.1, 1.5 MPa) for the signals received with 0.5 MHz transducer. The vertical axis represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.

sonation time (Figure 5.1). The integrated cavitation dose increases with PNP and was higher for smaller microbubbles for most of pressures compared to large and unfiltered MBs.

The influencing factors may be the stiffness of the MBs and surface tension which affects smaller microbubbles, while the larger MBs are inertia controlled. In addition, the number density is high for smaller MBs compared to larger bubbles which might produce more scattering and higher integrated cavitation dose.

To sum up, this future work results has shown that the size isolation is very important for understanding various MB behavior and their quantification that could measure



Figure 5.3: Integrated cavitation dose for DEF MBs at 30 cycle pulse duration and acoustic pressure of 0.5,1.1,1.5 MPa. PLots represents the cavitation dose for subharmonic $(0.5\pm0.2 \text{ MHz})$, ultraharmonic $(1.5\pm0.2 \text{ MHz})$, and harmonic $(2\pm0.2 \text{ MHz})$ for unfiltered, (blue) large (green) and small bubbles (red) respectively. The vertical axis represents the RMS FFT amplitude (arbitrary unit) and horizontal axis represents the time (s).

sonoporation outcomes at narrow size distribution. The conclusive results for MB acoustic behaviours at narrow size distribution could be achieved estimating the resonance bubble sizes for insonation frequency and isolate bubble sizes below and above that size. More experiments should be performed at same conditions to verify these results. Finally, future work should correlate the integrated cavitation dose with cell permeability and viability. Appendices

Appendix A

Time domain signal

The linear oscillation and nonlinear oscillation of the MBs are distinguished based on the frequency contents on the FFT plots. The linear oscillation exhibits fundamental (f)and second harmonics (2f) while the nonlinear oscillation exhibits subharmonic (f/2), ultraharmonic (3f/2, 5f/2) and higher harmonics.

In term of oscillation of MBs (time domain signals) it has been mentioned that the symmetric, regular and periodic oscillation give rise to linear oscillation while asymmetric, irregular and aperiodic oscillation results nonlinear oscillation of MBs. Our results demonstrated that there is a nonlinear regimes at 0.5 to 1.5 MPa where different patterns of the oscillation from MBs can be seen Figure 3.1 and 3.2. The portion of the TD signal at $30-40\mu$ s, for Figure 3.1 is shown in Figure A.1.

At 0.5 MPa, every μ s (e.g 31, 32, 33) repetitions of the one kind of oscillation pattern was seen while another kind of oscillation pattern was reported every μ s (e.g 31.5, 32.5, 33.5). The difference between two patterns was of 0.05 units in average toward the expansion while it was 0.11 units at contraction side which is an example of asymmetric oscillation. We define this as weak nonlinear oscillation where the fundamental, second



Figure A.1: Portion of the TD signal at $30-40\mu$ s, at a pressure of 0.5, 1.1 and 1.5 MPa shown by a, b, and c respectively. The vertical axis represent amplitude (Arbitrary unit) and horizontal axis represent time (s).

harmonic and third harmonics were emitted shown in Figure 3.1. At 1.1 MPa, repetition of the patterns of oscillation was seen every 2μ s (e.g 31.5, 33.5) with three small oscillation in a pattern, of lower amplitude in between which represents very strong nonlinear oscillation with an emissions of additional ultraharmonics, second and third harmonics of higher magnitudes. At 1.5 MPa the oscillation is very irregular, aperiodic and asymmetric which is difficult to predict its behaviour. This is called chaotic oscillation and its frequency content consists of broadband emissions.

Figure A.2 demonstrated that at 1.1 MPa, the MBs show expansions like behaviour where the average amplitude for expansion was greater by 0.12 units than the average



Figure A.2: Portion of the TD signal for 10μ s, at a pressure of 1.1 and 1.5 MPa shown by a, and b respectively. The vertical axis represent amplitude (Arbitrary unit) and horizontal axis represent time (s).

amplitude for contractions. At 1.5 MPa, the MBs show contraction like behaviour where the average amplitude for contraction was greater by 0.25 units than the average amplitude for expansion. This kind of behaviours are described by (62) following the bubbles buckling and ruptured regimes as described by (29). It was mentioned at low amplitude, the compression -only behaviour was attributed to a reversible buckling of the lipid shell while the expansion only behaviours are attributed to rupturing regimes.

Appendix B

Experiments Results

The PCD experiments results for ART and DEF MBs at PNP of 0, 1.1 and 1.5 MPa and 30 cycle pulse duration received with 2.25 MHz transducer are shown in Figures (B.1 and B.2). Similar results for ART and DEF MBs at PNP of 0, 1.1 and 1.5 MPa and 5 cycle pulse duration received with 2.25 MHz are shown in Figures (B.3 and B.4). In addition the spectrograms received with 0.5 MHz transducers for DEF and ART MBs for 5 cycle pulse duration are shown in Figure B.5. The FFT spectra from second set of experiment for both MBs are shown in Figure B.6. Moreover the results for cavitation dose from second set of experiments are shown in Figures (B.7,B.8, B.9 and B.10). These results have demonstrated similar behaviours of microbubbles that are discussed in result chapters and support the conclusion drawn in this projects.


Figure B.1: ART MB spectrograms as a function of PNP for signals received with 2.25 MHz receiver transducer. Results of two different experiments are shown for 0.5, 1.1, 1.5 MPa at 30 cycle pulse duration. The vertical axis on spectrogram represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.



Figure B.2: DEF MB spectrograms as a function of PNP for signals received with 2.25 MHz receiver transducer. Results of two different experiments are shown for 0.5, 1.1, 1.5 MPa at 30 cycle pulse duration. The vertical axis on spectrogram represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.



Figure B.3: ART MB spectrograms as a function of PNP for signals received with 2.25 MHz receiver transducer. Results of two different experiments are shown for 0.5, 1.1, 1.5 MPa at 5 cycle pulse duration. The vertical axis on spectrogram represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.



Figure B.4: DEF MB spectrograms as a function of PNP for signals received with 2.25 MHz receiver transducer. Results of two different experiments are shown for 0.5, 1.1, 1.5 MPa at 5 cycle pulse duration. The vertical axis on spectrogram represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.



Figure B.5: ART and DEF MBs spectrograms as a function of PNP for signals received with 0.5 MHz receiver transducer. Results of two different bubbles are shown for 0.5, 1.1, 1.5 MPa at 5 cycle pulse duration. The vertical axis on spectrogram represents frequency (Hz) and horizontal axis represents time (s). The color bar represents the frequency spectral amplitude, with red the highest and blue the lowest.



Figure B.6: ART and DEF MBs FFT spectra for 10ms insonation time from second set of experiment as a function of PNP for signals received with 2.25 MHz receiver transducers. Results of two different bubbles are shown for 0.5, 1.1, 1.5 MPa at 30 and 5 cycle pulse duration. The vertical axis represents amplitude (dB) and horizontal axis represents frequency (Hz).



Figure B.7: RMS FFT amplitude vs time plots for DEF MB at 30 cycles pulse duration from signals of experiment 2. (a), (b),(c) and (d) represent the cavitation dose for subharmonic $(0.5\pm0.2 \text{ MHz})$, ultraharmonic $(1.5\pm0.2 \text{ MHz})$, harmonic $(2\pm0.2 \text{ MHz})$ and ultraharmonic $(2.5\pm0.2 \text{ MHz})$ respectively. The vertical axis represents the RMS FFT amplitude (arbitrary unit) and horizontal axis represents the time (s).



Figure B.8: RMS FFT amplitude vs time plots for ART MB at 30 cycles pulse duration from signals of Experiment 2. (a), (b),(c) and (d) represent the cavitation dose for subharmonic $(0.5\pm0.2 \text{ MHz})$, ultraharmonic $(1.5\pm0.2 \text{ MHz})$, harmonic $(2\pm0.2 \text{ MHz})$ and ultraharmonic $(2.5\pm0.2 \text{ MHz})$ respectively. The vertical axis represents the RMS FFT amplitude (arbitrary unit) and horizontal axis represents the time (s).



Figure B.9: RMS FFT amplitude vs time plots for DEF MB at 5 cycles pulse duration. (a), (b),(c) and (d) represent the cavitation dose for subharmonic $(0.5\pm0.2 \text{ MHz})$, ultraharmonic $(1.5\pm0.2 \text{ MHz})$, harmonic $(2\pm0.2 \text{ MHz})$ and ultraharmonic $(2.5\pm0.2 \text{ MHz})$ respectively. The vertical axis represents the RMS FFT amplitude (arbitrary unit) and horizontal axis represents the time (s).



Figure B.10: RMS FFT amplitude vs time plots for ART MB at 5 cycles pulse duration. (a), (b),(c) and (d) represent the cavitation dose for subharmonic $(0.5\pm0.2 \text{ MHz})$, ultraharmonic $(1.5\pm0.2 \text{ MHz})$, harmonic $(2\pm0.2 \text{ MHz})$ and ultraharmonic $(2.5\pm0.2 \text{ MHz})$ respectively. The vertical axis represents the RMS FFT amplitude (arbitrary unit) and horizontal axis represents the time (s).

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